IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

U.S. PATENT APPLICATION

FOR:

DEVICE AND METHOD FOR CYTOLOGY SLIDE PREPARATION

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FIELD OF INVENTION

The present invention relates generally to the preparation of cytology slides and, more particularly, to a device and associated method for preparing cytology slides.

BACKGROUND OF THE INVENTION

Cytology slides are prepared to screen and diagnose cellular samples taken from, for example, samples from the uterine cervix, urine, sputum, blood, fine needle aspiration biopsy, urethral, bronchial brushings and washings, cerebral spinal fluid, and other body fluids. The reliability and efficacy of the screening methods of these slides are measured by their ability to diagnose infections, precancerous lesions or cancerous lesions while at the same time avoiding false positive or negative diagnosis. The reliability of these slides is a primary issue. Often, the results are not accurate or are unreadable. Thus, there is a constant effort to improve the reliability and efficacy in the preparation of cytology samples.

One of the most common uses of cytology slides is for screening and diagnosis of a cervical sample. Carcinoma of the cervix is one of the most common malignancies in women, causing nearly 5,000 deaths per year in the United States. Approximately 60% of these cases are associated with absent or deficient screening. Approximately 25% of the screening

failures are the result of errors in cervical sampling or smear interpretation.[1]

Screening for precancerous or cancerous changes of the uterine cervix traditionally involves microscopic assessment of cervical Papanicolaou smears, called Pap smears. This traditional method for screening requires scraping a woman's cervix with a sampling device, such as a cotton applicator stick, spatula or brush, and smearing this sample onto a slide for review by a medical lab professional. The specimen is gently spread across a slide to evenly distribute the cell sample. The slide is then fixed, stained, and examined under a light microscope for cellular abnormalities.

In carrying out this operation, the portion of the cell sample that is smeared onto the slide may contain blood, mucus, inflammatory cells, and clumps of cells. Accurate interpretation of up to 40% of conventional Pap smears are compromised by the presence of blood, mucous, obscuring inflammatory cells, scant cellular material and air-drying artifacts.[2] The presence of these contaminants can obscure many of the cells, causing important precancerous lesions to be missed when the slide is reviewed at the lab or, alternatively, making the entire slide unreadable.

One of the problems with the conventional Pap smear is how quickly the sample dries out once it is smeared on the slide.

With a conventional Pap smear, the sample must be fixed immediately in order to avoid the drying out of the cells, which ruins the sample.

Another problem with the conventional Pap smear is the frequent inaccuracy of the test result. Common inaccuracies include both false positive and false negative Pap test results. A false positive Pap test occurs when a patient is told she has abnormal cells when the cells are actually normal. A false positive result may require a woman to undergo unnecessary and costly medical procedures. A false negative Pap test result occurs when a specimen is called normal, but the woman has a lesion. A false negative Pap test may delay the diagnosis and treatment of a precancerous or even a cancerous condition.

The conventional Pap smear has false negative rates ranging from 10-50%, with up to 90% of those false negatives due to limitations of sampling or slide preparation.[3] To decrease false negative rates associated with interpretation error, re-screening a portion of the negative smear or recalling the patient for another sample is required.

Concern over the frequency of false-negative results of the traditional Pap smear has led to the development of a variety of other technologies or clinical strategies, such as liquid-based cytology systems, to improve Pap testing. For example, the Cytyc, Inc. (Marlborough, M.A.), ThinPrep® and the TriPath, Inc.

(Burlington, N.C.), CytoRich® Pap test systems are two commercially available, FDA approved fluid-based methods used for the collection and preparation of cervicovaginal samples.

With the ThinPrep® system, a gynecologic sample is collected in the same manner as the conventional Pap test using a broom-type device or plastic spatula and endocervical brush combination, but rather than smearing the cytological sample directly onto a microscope slide, this method suspends the sample cells in a fixative solution (i.e. PreservCyt®). The ThinPrep® slide preparation system uses an automated apparatus called a Cytyc 2000® that involves filtration using vacuum pressure and positive pressure-transfer steps to prepare cytology slides.[4]

With the CytoRich® slide preparation system, the gynecologic sample is also collected in the same manner as the conventional Pap test. Like the ThinPrep® system, the CytoRich® system also places the sample in a liquid medium for further purification prior to analysis. CytoRich® specimens are processed using two centrifugation steps through a gradient solution to separate the diagnostic cells from the interferring material. The cells are ultimately re-suspended in a final preparation that is applied to the slide using a special pipetting apparatus (Autocyte Prep System®) provided by the manufacturers (Tripath, Inc.). This transfer step can also be

performed manually. Thereafter, a sample is placed on a slide and analyzed by cytology.

These new methods have demonstrated increased quality in the preparation of the sample, improved detection rates, and a reduced need for patients who must return for repeat smears. However, in both the ThinPrep® and the CytoRich® slide preparation systems, a time consuming and expensive procedure is followed to prepare a mono-dispersed layer of cells on a cytology slide.

Although each of these systems reduces significantly the false-negative rate of the traditional Pap smears, the health care market has been slow in adopting these systems because of their cost and preparation time compared to the conventional Pap smears.

SUMMARY OF THE INVENTION

The invention provides a device and a method of using the device that overcomes these problems. The device and associated method provide a better quality prepared cytology slide having a more even distribution of cells with less interference from debris than the conventional Pap method, while offering an easier, quicker and more economical procedure as compared to known liquid-based medium methods. Thus the device and associated method facilitate the preparation and screening process of cytology slides.

According to one embodiment, a device for facilitating the preparation of cytology slides comprises a first cover having an inside surface, a second cover having an inside surface and an interposed spine. The spine may be replaced by one or more hinges. The first and second cover are pivotably secured to the spine so as to be foldable into a book form capable of an open and closed position. An absorbent material is mounted on the inside surface of the first cover and a filter overlays the absorbent material. The cytology slide is removeably mounted to the inside surface of the second cover. The slide is positioned on the inside surface of the second cover to contact with the filter when the book-like device is in the closed position.

In another aspect, the invention is directed to a method of preparing cytology slides. The method comprises combining a cellular sample with a liquid-based medium to create a solution, mixing the solution, extracting the sample or an aliquot of the sample from the solution and providing a slide preparation device. The device comprises a first cover having an inside surface, a second cover having an inside surface and an interposed spine. The first and second cover are pivotably secured to the spine so as to be foldable into a book form. An absorbent material is mounted to the inside surface of the first cover and a filter overlays the absorbent material. A slide is removeably mounted to the inside of the second cover so that the

filter contacts the slide when the device is in the closed position. The method further comprises applying the sample to the filter, closing the book form so that the filter containing the sample contacts the slide, and applying a pressure to the contact surface to effectively transfer the specimen to the slide. The range of pressures appropriate for the absorbent matter and the filter in the device is provided by the design of the device. In this way, a cytology slide containing an evenly distributed number of cells is prepared quickly and reliably.

In another aspect of the invention, the invention is directed to a device for facilitating the preparation of a plurality of cytology slides at the same time. The device comprises a cover having an inside surface, a base having an inside surface, and an interposed hinge. The cover and base pivotably secure to the hinge so as to be foldable into a book form capable of an open and closed position. A plurality of absorbent material are mounted on the inside surface of the cover and a filter overlays the absorbent material. A plurality of cytology slides corresponding to one of the absorbent material are mounted to the inside surface of the base. Each slide is positioned on the inside surface of the base to contact the corresponding absorbent material and the filter when the book form is in the closed position

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1A is an exploded view of one embodiment of the devices;

Fig. 1B is an isometric view of one embodiment of the device;

Fig. 1C is an isometric view of one embodiment of the device;

Fig. 2A is an isometric view of another embodiment of the device illustrating a single continuous hinge;

Fig. 2B is an isometric view of one embodiment of the device illustrating a hinge having a number of interlocking pieces 34;

Fig. 3 is an isometric view of another embodiment of the device illustrating a pocket 46 and fastening means 60;

Fig. 4A is a partial view of the front cover of one embodiment of the device illustrating the recess or well 36;

Fig. 4B is a partial view of the front cover of one embodiment of the device illustrating the absorbent material and filter extending above the inner surface; and

Fig. 5 is a partial view of the front cover of one embodiment of the device illustrating a plurality of supports 50.

Fig. 6A illustrates the cell distribution of a cervical sample using the device and method of the present invention.

Fig. 6B illustrates the cell distribution of a conventional Pap smear (archival)at a magnification of x120.

Fig. 7 illustrates an exploded view of an embodiment of the device capable of preparing multiple cytology slides at the same time.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

Hereinafter, the invention will be described in greater detail with reference to the preferred embodiments. However, it is understood that the device and associated method of the invention are applicable for the preparation of cytology slides for any cellular or tissue samples, preferably clinical cellular samples, known in the art.

Referring now to Fig. 1A, in one embodiment of the invention, a kit 100 for facilitating the preparation of cellular samples includes a device 10 comprising a front cover 12, a rear cover 16, and a spine 24 connecting the front and rear covers 12, 16. The spine 24 spaces the front cover 12 and the back cover 16 when folded in the closed position and creates a book-like form, wherein the inside 14 of the front cover 12 and the inside 18 of the back cover 16 face and contact against each other. In an alternative embodiment illustrated in Fig.

1A, the front cover 12 does not need to be the same size as the back cover 16.

The covers 12, 16 and spine 24 may be made of a single piece of flexible material. Such flexible material can include, for example, cardboard or some flexible plastics to which a controlled pressure may be applied to the outside covers 12, 16 without puncturing or cracking the covers. The material should not absorb the sample or solution applied to the filter 22 and absorbent material 20 (described in more detail below). The material should also be selected to enable various users, such as, for example, lab technicians and doctors, to apply consistent or controlled amount of pressure to prevent over squeezing the solution, resulting in spillage or distortion of the morphology of the sample by the exertion of too much pressure.

Alternatively, the device 10 can be made from an inflexible material. Such inflexible material can include, for example, wood, wood-derived materials, metals or inflexible plastics. When inflexible materials are used for the front and back covers, it may be preferable to use one or more hinges to connect the front and back covers. In the embodiment illustrated in Figs. 2A and 2B, a hinge 30 connects the front cover 12 and back cover 16. The hinge 30 is constructed so that the front cover 12 and back cover 16 face and contact each other

when the book form is in the closed position. The hinge 30 can be one single, continuous piece 32 (Fig. 2A) or, alternatively, a number of flexible or interlocking pieces 34 (Fig. 2B).

In Fig. 1B, the device is shown in the open position. Front cover 12 is flipped open, exposing an absorbent material 20 and a filter 22 attached to the inside 14 of the first cover 12. A slide 40 is also shown mounted to the inside 18 of the back cover 16 of the device 10.

The absorbent material 20 is hydrophilic in nature. In particular, the absorbent material is absorbent to both water and alcohol-based solutions and is preferably uniform in its absorbency. The absorbent material is preferably inert and stable so as not to react with the alcohol solution or components of the cellular sample. The absorbent material preferably possesses stability properties enabling the finished device to be stored for several years. Standard filter papers or blotting papers may be used as an absorbent material. For example, one suitable absorbent material is that sold by Bio-Rad® (Hercules, California) called Absorbent Filter Paper (Extra Thick) consisting of 100% cotton fiber. Alternatively, the absorbent material can be 100% double-sided, twill-pattern woven cotton (e.g. Wipers Natural; TX306 made by TexWipe® Company), or polyvinyl acetal foam (HydroSponge™; TX5390 made by Tex Wipe® Company), Gelman's® Absorbent Pad, Cellulose Fiber Filter Pad,

Whatman's® Glass Microfibre filters, Whatman's® cellulose paper, TexWipe® Company (Upper Saddle River, NJ) trademarked Absorbond® (Wipers Synthetic; TX404) consisting of 100% hydro-entangled polyester, Denville® Scientific Hyblot Mater, Millipore® cellulose absorbent pads or any other absorbent material or combination of materials known in the art.

The absorbent material 20 adheres to the inside 14 of the front cover 12 by, for example, an adhesive or, alternatively, by a heat spot weld. It should be understood that the adhesive is selected so as not to interfere with the properties of the absorbent material, as mentioned above. One such suitable adhesive is gel cyanoacrylate sold by Loctite under the trademark Loctite 4541®. Alternatively, the adhesive can be hot melt or acrylic or synthetic rubber based adhesives or any adhesive known in the art.

The filter 22 overlays the absorbent material 20. Any filter known in the art can be used. The filter 22 is preferably a polycarbonate film containing approximately 3 to 8 micrometer pores. The pore sizes can be varied to enable the filter to capture the desired cells on the surface of the filter while allowing the debris to pass through to the absorbent material 20. The filter should also be inert, stable, and alcohol resistant. Alternatively, the filter can be nylon,

cellulose, polyester, teflon, polytetrafluorethylene or any other filter material known in the art.

An adhesive to attach the filter 22 to the absorbent material 20 is selected so as not to interfere with the sample placed on the filter 20. Revertex Chemicals Ltd. (Malaysia) sells some suitable adhesives for attaching the filter 22 to the absorbent material 20, such as, for example, starch, dextrin, latex and casein. Alternatively, the adhesive can be ethylene vinyl acetate and polyvinyl acetate emulsion adhesive for lamination of the filter 22 and the absorbent material 20. In other embodiments, the filter 22 can be held in proximity to the absorbent material 20 by mechanical means, such as, for example, a rim of plastic to hold the overlapping filter 22 onto the absorbent material 20 disk.

The slide 40 is a standard slide, made of either glass or plastic, commonly used in cytology. In the embodiment of Fig. 1A, the slide 40 is mounted to the inside 18 of the back cover 16 in such a way that enables the slide 40 to be easily removed. In the embodiment of Fig. 1B, the slide 40 is mounted to the inside 18 of the back cover 18 by a piece of adhesive or tape 42. However, any commonly known method known in the art that accomplishes this goal can be used to mount the slide. For example, the slide 40 can be press-fit into a cutout portion 44 of the inside 18 of the back cover 16, held by Velcro® strap, or

placed in a pocket 46 formed in the inside 18 of the back cover 16 (Fig. 3) to facilitate removal of the slide 40.

In another embodiment, the inside 18 of the back cover 16 includes finger recesses 28 to aid a user in lifting the slide 40 out of the back cover (Fig. 1B and 1C). The finger recesses 28 should be positioned away from the preparation area to avoid contamination of the transferred sample.

The absorbent material 20 with the attached filter 22 are positioned on the inside 14 of the front cover 12 of the book-like device 10 so that when the book-like device 10 is closed, the filter 22 contacts a central location on the slide 40.

In another embodiment, as illustrated in Fig. 4A and 4B, the absorbent material is placed in a recess or shallow well 36 formed in the inside 14 of the front cover 12 so that the upper portion of the absorbent material 20 having the overlaying filter 22 (as will be described in more detail below) extends above the inside surface 14 of the front cover 12. This recess or shallow well 36 provides the added benefit of holding any excess solution that escapes from the bottom of the absorbent material 20. In yet another embodiment, the absorbent material 20 is held in place on the inside 14 of the front cover 12 by a plurality of supports 50, as illustrated in Fig. 5.

When the book-form is closed, the inside 14 of the front cover 12 and the inside 16 of the back cover 10 contact each

other. In the embodiment in which the upper portion of the absorbent material 20 and filter 22 extend above the inside surface 14 of the front cover 12, there is no need to exert an additional pressure to transfer the sample to the slide 40 from the filter 22, when the back cover 10 comes into contact with front cover 12. The slide 40 , which can be flush with the inside 18 of the back cover 16 will compress the absorbent material 20 and filter 22, thus transferring the sample to the slide 40.

As illustrated in Fig. 3, the kit 100 may optionally include a mechanical fastening means 60 to lock together and maintain the contact of the inside surfaces 14, 18 of the front and back covers 12, 16, respectively, when the book-form device 10 is closed. Any mechanical fastening means known in the art, such as Velcro®, tape, clasps, snaps, or the like, can be used. When the book-form device 10 is closed, causing the filter 22 to contact the slide 40, the appropriate amount of controlled pressure may be applied by fastening the fastening means 60. In an embodiment including a clasp or snap fastening means 60, an audible snap is preferably heard when the fastening means is fastened. This audible snap informs the user that a controlled amount of pressure is being applied to transfer the sample from the filter 22 to the slide 40.

A cellular sample is any cellular or tissue specimen collected from a subject for the purposes of screening or diagnosing, commonly known in the art, that is capable of being suspended in a liquid-based medium. Such samples are obtained, for example, from the cervix, the mouth or throat, urine sediments, lymph nodes, esophagus, bronchial, breast (i.e. nipple discharge), skin lesions, thyroid, blood and disruptive tissue biopsies.

In the present invention, the kit 100 is used to prepare a cytology slide with a mono-dispersed cell sample. In the broader aspects of the invention, there is no limitation on the collection and handling of samples as long as consistency is maintained. The cell or tissue sample is obtained by methods known in the art, such as, biopsies, surgical resections, standard bodily fluid collection techniques or the like.

In an embodiment in which the cellular sample is a cervical smear, the sample is taken from the cervix. The present invention employs standard techniques for collecting cervical cells; for example, it generally involves the insertion of a cervical brush or broom device and the taking of cells from the surface of the cervix and the endocervix.

The cellular sample is preferably placed in any liquid-based cytology medium used and known in the art. One example of such a cytology medium is known as a Universal Collection Medium

(UCM) from Digene Corp. (Gaithersburg, MD). UCM is a cell collection medium which preserves both cell morphology and cellular biomolecules for quantitative analysis in a cell sample so that multiple assays can be carried out from a single patient sample.[5] Other liquid-based cytology media can be used, such as PreservCyt® from Cytyc, Inc. (Boxborough, MA) and CytoRich® Preservative Fluid from TriPath, Inc. (Burlington, N.C.).

One preferred method places the cellular sample in 1 milliliter of UCM. This solution is then suspended, by mixing. Mixing can be conveniently achieved, for example by vortexing the solution for approximately five to ten seconds.

After suspending the cells, a sample is removed carefully, usually with a pipette, and placed on the filter 22. If one milliliter of UCM is used, it may be preferable to use an aliquot of approximately 200 microliters from the solution. This sample is then applied to the surface of the filter 22. One skilled in the art understands that other liquid media may be used and the sample may be suspended in varying amounts of such media. The solution and other debris pass through the filter 22 to the absorbent material 20 while the cells are captured on the surface of the filter 22. The book-like device 10 is then closed so that the filter 22 contacts the slide 40 for about 15 seconds while a moderate pressure, created when the covers are closed and locked together, is applied to the front

and back covers 12, 16, respectively. The device or user applies from 200 to 1000 grams of pressure to the slide and specimen for optimum transfer of the cytology specimen to the slide.

The pressure applied to the covers 12, 16 transfers the cells captured by the filter 22 to the slide 40. After the pressure is applied, the slide 40 can be removed from the book-like device 12 and fixed and stained in the routine manner.

In order to optimally preserve the cell detail and maintain the morphologic characteristics, without distortion, the cells are preferably wet-fixed before any air drying occurs.

Conventional methods known in the art can be used for fixing, staining and analyzing the sample. The following provides a general outline of such conventional techniques but should not be construed to limit the applicability of the invention to the alternative cytology preparation techniques known in the art.

Most laboratories use a 95 percent ethanol solution for most routine preparations that require fixations. However, other fixatives known in the art can be used. Most of these fixatives are composed of polyethylene glycol in an alcohol base and are applied either from a dropper bottle or in spray form. Coating fixatives are also widely used for cervical smears.

These fixatives cover the sample in a waxy coating that protects it from damage.

One advantage of using the absorbent material 22 is that the cell sample will not dry out as quickly. When the book-like device 10 is closed, the absorbent material 22 keeps the sample moist as it is transferred to the slide since a liquid-based solution is used and the absorbent material 20 is held in close physical proximity to the specimen. The device 10 maintains a "moist" or "humid" environment so that the cells can be preserved in the device 10 before fixing for up to 30 minutes following application on the slide 40. Once on the slide 40, the cell sample should be preserved as mentioned above.

After the cells are transferred to the slide and the cells are fixed, the cellular sample is generally stained. Staining may be required to retain the transparency of the cells and display variations of cellular maturity and metabolic activity. Any stain or staining method known in the art can be used.[6] The universal stain for cytological preparations is the Papanicolaou stain. Alternatively, any stain known in the art, such as Hematonylin and Eosin stain or special chemical stains or immunochemical stains, or in-situ hybridization can be used.

Once any staining technique has been completed, the slide may be mounted in a suitable medium satisfactory for microscopy. There are many commercially available mountants that can be used

that are well known in the art. These mountants protect the sample from dust and damage and should harden rapidly and not discolor or crystallize during long-term storage.

A coverslip may then be placed over the sample. Coverslipping has traditionally been achieved using a thin glass slip (less than 0.17 millimeters) or continuous film. Various sizes are commercially available but the entire sample preparation should be covered by the mountant and the coverslip.

Referring now to Fig. 7, in another embodiment of the invention, a kit 200 for facilitating the preparation of a plurality of cellular samples includes a cover 210, a base 230 and at least one hinge 205 for closing cover 210 over base 230 to create a book-like form.

The cover 210 and base 230 may be made of a single piece of flexible material. Such flexible material can include, for example, cardboard or some flexible plastics to which a controlled pressure may be applied to the outside surface of the cover without puncturing or cracking the cover. The material should not absorb the sample or solution applied to the filters and absorbent material, as described in more detail above. The material should also be selected to enable various users, such as, for example, lab technicians and doctors, to apply consistent or controlled pressure to prevent over squeezing the

solution, resulting in spillage or distortion of the morphology of the sample by the exertion of too much pressure.

Alternatively, cover 210 and base 230 can be made from an inflexible material. Such inflexible material can include, for example, wood, wood-derived materials, metals or inflexible plastics. In one embodiment, the weight of the material used for the cover provides enough pressure to effectively transfer the sample from the absorbent material and filter onto the cytology slide. In experimenting with the appropriate weight to be applied to effectively transfer the sample, over one hundred slides were prepared using the method discussed above and weights up to 1200 grams were applied to the cover 210. The slides were then fixed, stained and evaluated. Based on the morphological evaluation, it was determined that the cell morphology was altered, e.g. the nuclei were slightly enlarged, when weights close to 1200 grams were used. It was also determined that the used of weights of less than 300 grams did not effectively transfer the samples from the absorbent material and filter to the slides.

In an alternative embodiment, clamps can be used to close cover 210 and base 230 so that the samples may be effectively transferred.

The cover 210 supports a strip 212 holding a plurality of pieces of absorbent material 20 and a filter strip 22 overlaying

the absorbent material 20. The absorbent material 20 and filter 22 are made from the same materials discussed above. In alternate embodiments, separate filters corresponding to each piece of absorbent material 20 held by the strip 212 or, alternatively, a filter 22 overlaying a number of pieces of absorbent material 20 can be used.

The upper portion of the absorbent material 20 and filter 22 extend above the top surface of strip 212 so that when cover 210 is closed over base 230, the samples applied to the absorbent material 20 and filter 22 are effectively transferred to the corresponding slides 40 held by slide carrier 232.

The strip 212 holding absorbent material 20 and filter 22 is supported by cover 210 in a channel 214 formed in the inside surface 216 of the cover 210. In one embodiment, strip 212 is press fit into the channel 214. In another embodiment, strip 212 includes feet 220 projecting from the bottom side of strip 212 which are press fit into recesses 222 formed in the bottom surface 213 of channel 214. The feet 220 also serve to stabilize and position strip 212 in relation to cover 210 and keep the strip 212 from sliding out of the channel 210. In another embodiment, strip 212 can be slid into and resiliently held by channel 214.

The strip 212 can also include positioning members 226 projecting from the sides of strip 212. Positioning members 226

are received by recesses 227 formed in the sides of channel 214 to ensure that strip 212 is properly positioned in channel 214.

The absorbent material 20 is adhered to the top surface of strip 212 by, for example, an adhesive or, alternatively, by a heat spot weld. It should be understood that the adhesive is selected so as not to interfere with the properties of the absorbent material, as mentioned above. Alternatively, the adhesive can be hot melt or acrylic or synthetic rubber based adhesives or any adhesive known in the art. The filter overlays the adhesive pad in the same manner as discussed above.

The base 230 supports a slide carrier 232 capable of holding a plurality of slides 40. The number of slides 40 held by slide carrier 232 corresponds to the number of pieces of absorbent material 20 on strip 212 supported by cover 210.

The slide carrier 232 containing slides 40 is supported by base 230 in a channel 234 formed in the top surface of base 230. In one embodiment, slide carrier 232 is press fit into channel 234. In another embodiment, slide carrier 232 includes feet (not shown) projecting from the bottom side of slide carrier 232. The feet are press fit into recesses 238 formed in the bottom surface of channel 234. The slide carrier 232 can also include positioning members, similar to positioning members 236 on strip 212, projecting from the sides of slide carrier 232 to ensure that slide carrier 232 is properly positioned in channel 234.

The slide carrier 232 includes a plurality of slots 240 separated by ridges or dividers 242 for securely holding slides 40 in place when the kit 200 is opened and closed. While slide carrier 232 is held in channel 234 in base 230, the top surface 244 of dividers 242 is flush with the top surface of base 230 to ensure proper closing of kit 200. Finger recesses 250 may be provided to facilitate the insertion and removal of the slides 40 from slide carrier 232.

In an alternate embodiment, the slide carrier 232 can be hingeably attached to base 230. In another embodiment, the slides 40 are inserted into separate, individual slots formed in the base 230 itself.

The method of applying or smearing a sample to any of the absorbent materials 20 and filter 22 and transferring the sample to the slide 40 is identical to the method described above for the book-like device 10. With the kit 200, one lab technician or doctor can prepare multiple slides at the same time.

Once the samples are transferred and the slides are prepared, the slides may be individually analyzed or stored in slide carrier 232 for future analysis.

EXAMPLE: COMPARISON WITH CONVENTIONAL PAP SMEAR

A cervical sample was taken from a female patient and placed in a one milliliter of Universal Collection Medium.[7] An aliquot of 200 microliters is removed from the solution and

applied to the surface of the filter 22 of the book-form device 10. The book-form device 10 was closed for approximately 15 seconds while a controlled amount of pressure was applied. The slide 40 was then removed from the book-form device 10 and the sample was fixed, stained and placed under a light microscope. Fig. 6A shows the morphology of cervical cells prepared by the method of the present invention.

Fig 6B was taken from archived routine Pap smears to show the morphology, distribution of cells and staining of stored samples fixed to slides for comparison. The morphology of the cells prepared by the method of the present invention shows better quality and more evenly distributed cells with less interference from debris than the conventional Pap smear.

Table 1, below, shows a comparison of Pap slides from the same patients made with the conventional Pap smear technique and the UCM book-form device Pap slide technique. These results demonstrate complete agreement between the two procedures.

Table 1. UCM vs. Conventional Pap*

Case ID	Conventional Pap	UCM Manual Pap
1W	WNL	WNL
2W	LSIL	HSIL
3W	WNL	WNL
4W	WNL	WNL
5W	WNL	WNL
6W	WNL	WNL
7W	WNL	WNL

8W	WNL	WNL
9W	WNL	WNL
10W	WNL	WNL
11W	WNL	WNL
12W	WNL	WNL
13W	WNL	WNL
14W	WNL	WNL
15W	WNL	WNL

100% Agreement

Key:
WNL - Within Normal Limits
LSIL - Low-grade Squamous Intraepithelial Lesions
HSIL - High-grade Squamous Intraepithelial Lesions
ASCUS - Atypical Squamous Cells of Undetermined Significance

It will be apparent to those skilled in the art that various modifications and variations can be made in the device and method of the present invention without departing from the spirit or scope of the invention. Thus, it is intended that the present invention embraces all such modifications and variations within the spirit and scope of the appended claims.

All articles, patents or other references cited or referred to herein are hereby incorporated herein in toto by reference.

References

[1] Sawaya, George F. (M.D.), Grimes, David A. (M.D.), "New Technologies in Cervical Cytology Screening: A Word Of Caution", Obstetrics and Gynecology, 1999, Vol. 94, pg.1, which is incorporated herein by reference.

[2] Davey DD, Nielsen ML, Rosenstock W, Tilde TS. "Terminology and specimen adequacy in cervicovaginal cytology: The College of American Pathologists interlaboratory comparison program experience", *Arch. Path. Lab. Med.* 1992, vol. 116, pgs. 903-907 which is incorporated herein by reference.

[3] van der Graaf Y, Vooijs GP, Gaillard HLJ, Go DMDS, "Screening errors in cervical cytological screenings", *Acta Cytologica,* 1987, vol. 31, pgs. 434-8; Dehner LP, "Cervicovaginal cytology, false-negative results, and standards of practice", *Am. J. Clin. Pathol*, 1993, vol. 99, pgs. 45-47; Gay JD, Donaldson LD, Goetliner JR., "False negative results in cervical cytologic studies", *Acta Cytologica,* 1985, vol. 29, pgs. 1043-1046; and Morell, ND, Taylor JR, Snyder RN, Ziel HK, Saltz A, Willis S., "False-negative cytology rates in patients in whom invasive cerviocal cancer subsequently developed", *Obstet. Gynecol.*, 1982, vol. 60, pgs. 41-45 which are incorporated herein by reference.

[4] Further information about the operation and efficiency of the ThinPrep® method is set forth in Obstetrics & Gynecology, Vol. 90, No. 2, pps. 278-284 in an article by Lee, Ashfaq, Birdsong, Corkill, McIntosh and Inhorn titled "Comparison of Conventional Papanicolaou Smears and a Fluid-Based, Thin-Layer System for Cervical Cancer Screening" which is incorporated herein by reference.

[5] Further information about the UCM is set forth in PCT International Application No. PCT/US98/26342 (European patent application number 98962066.1), filed on December 11, 1998 by Attila T. Lorincz and Yanlin Tang for a Universal Collection Medium which is incorporated herein by reference.

[6] Further information about fixation and staining methods and materials used is provided by the Health Care Financing Administration and set forth in "The Tutorial of Cytology" by Wied, Keebler, Koss and Reagan (1998, pps. 580-583), and in "Comprehensive Cytopathology" by Bibbo, (1991, pps. 882-892) which are incorporated herein by reference.

[7] Further information about the UCM is set forth in PCT International Application No. PCT/US98/26342 (European patent application number

98962066.1), filed on December 11, 1998 by Attila T. Lorincz and Yanlin Tang for a Universal Collection Medium which is incorporated herein by reference.